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# Direct RNA sequencing with modifications

- Jannes Spangenberg

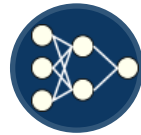


## Presentation outline



**Oxford Nanopore Technologies direct RNA sequencing**

- **Challenges and problems**



**RNA modification prediction using neural networks**

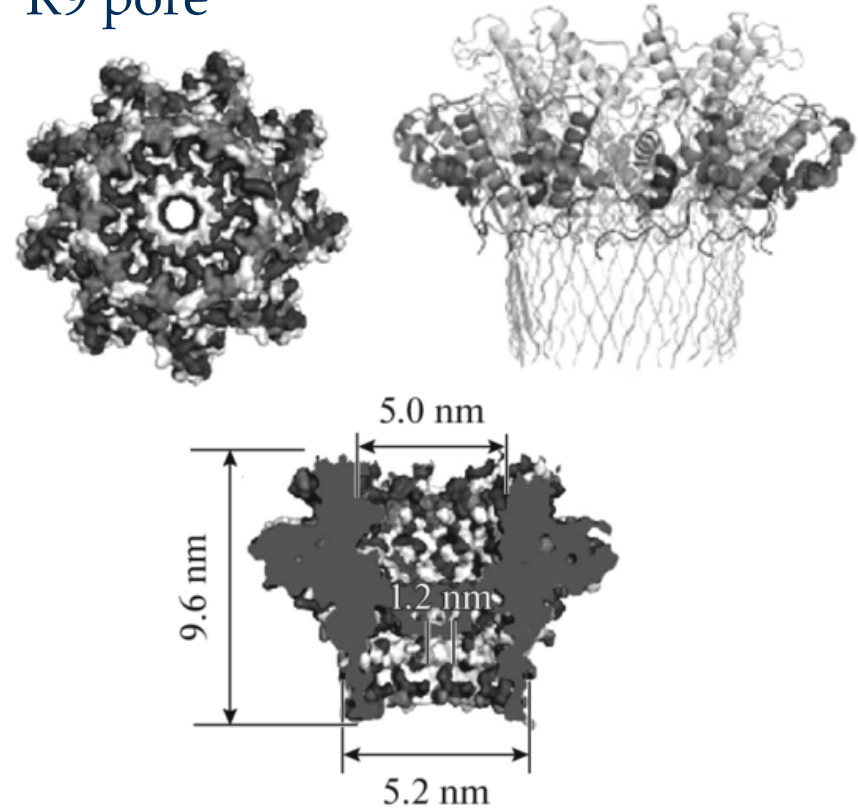


**Current projects and ideas**

## R9 pore

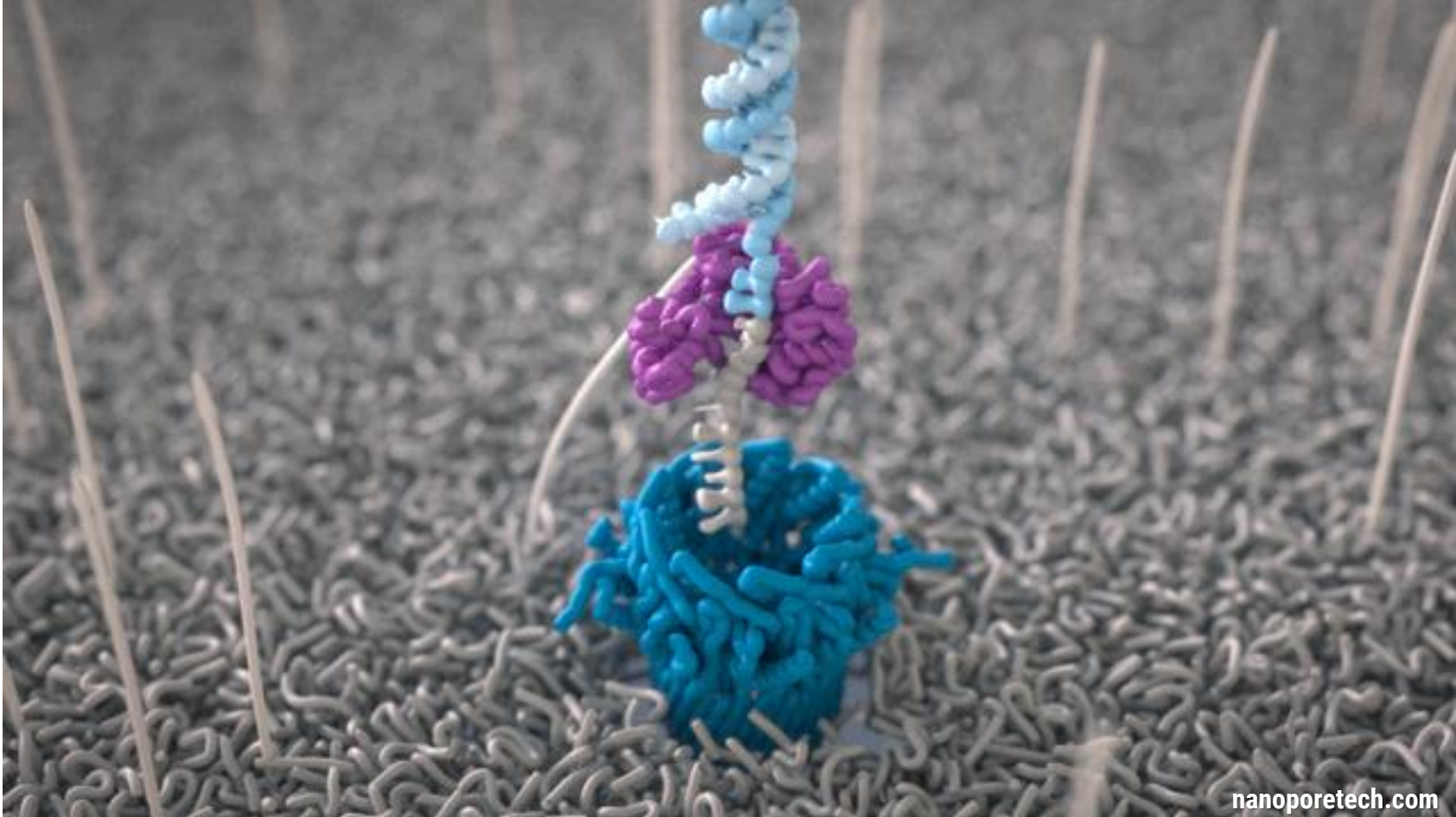
### What is a nanopore?

- Nanometer-sized pore made up of proteins
- Based on bacterial membrane pore complexes
- Improved by deliberate mutation to measure nucleotides
- Pore resides in a membrane



Barkova, D.V., Andrianova, M.S., Komarova, N.V. *et al.* Channel and Motor Proteins for Translocation of Nucleic Acids in Nanopore Sequencing. *Moscow Univ. Chem. Bull.* 75, 149–161 (2020).

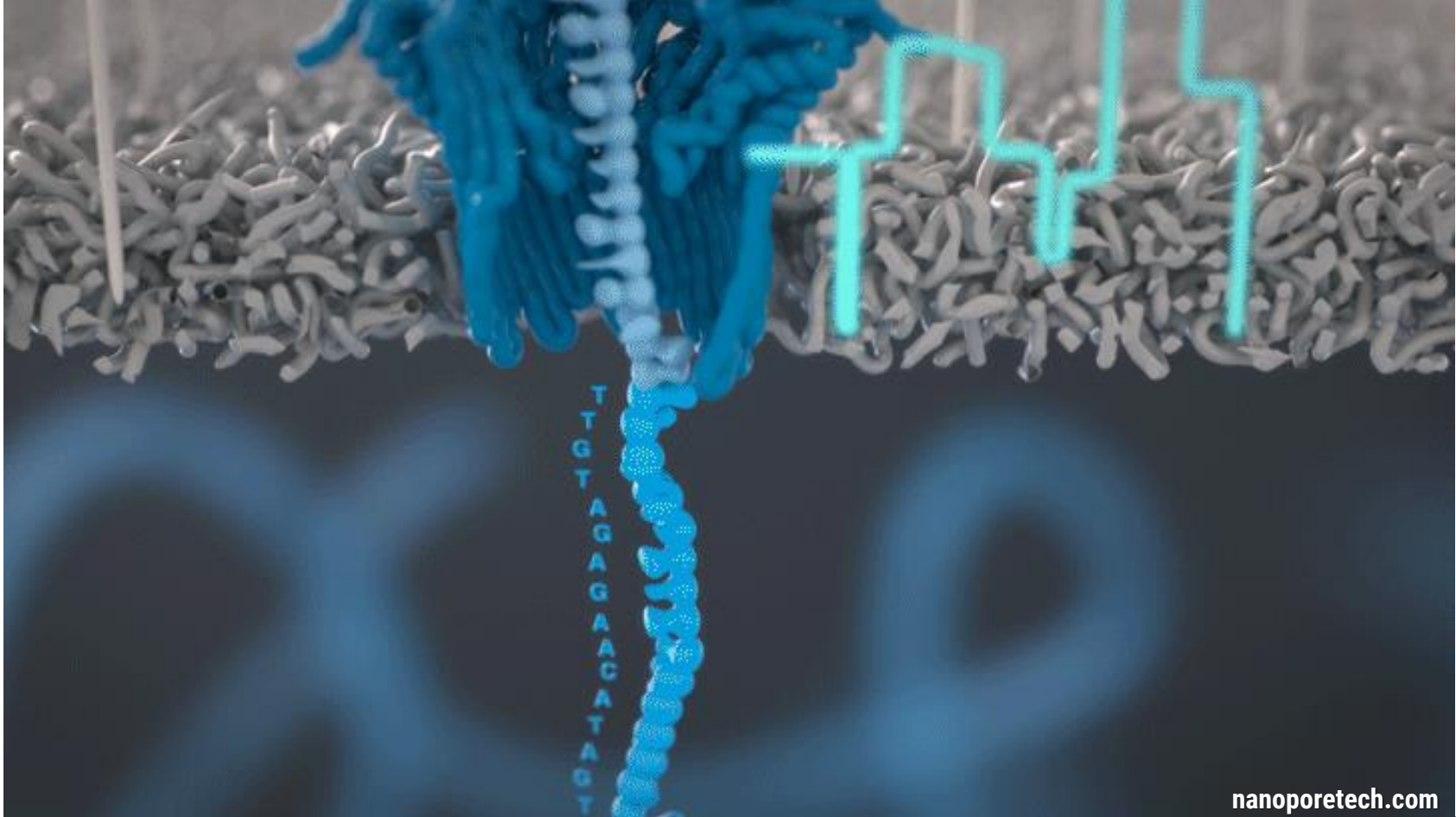




[nanoporetech.com](http://nanoporetech.com)



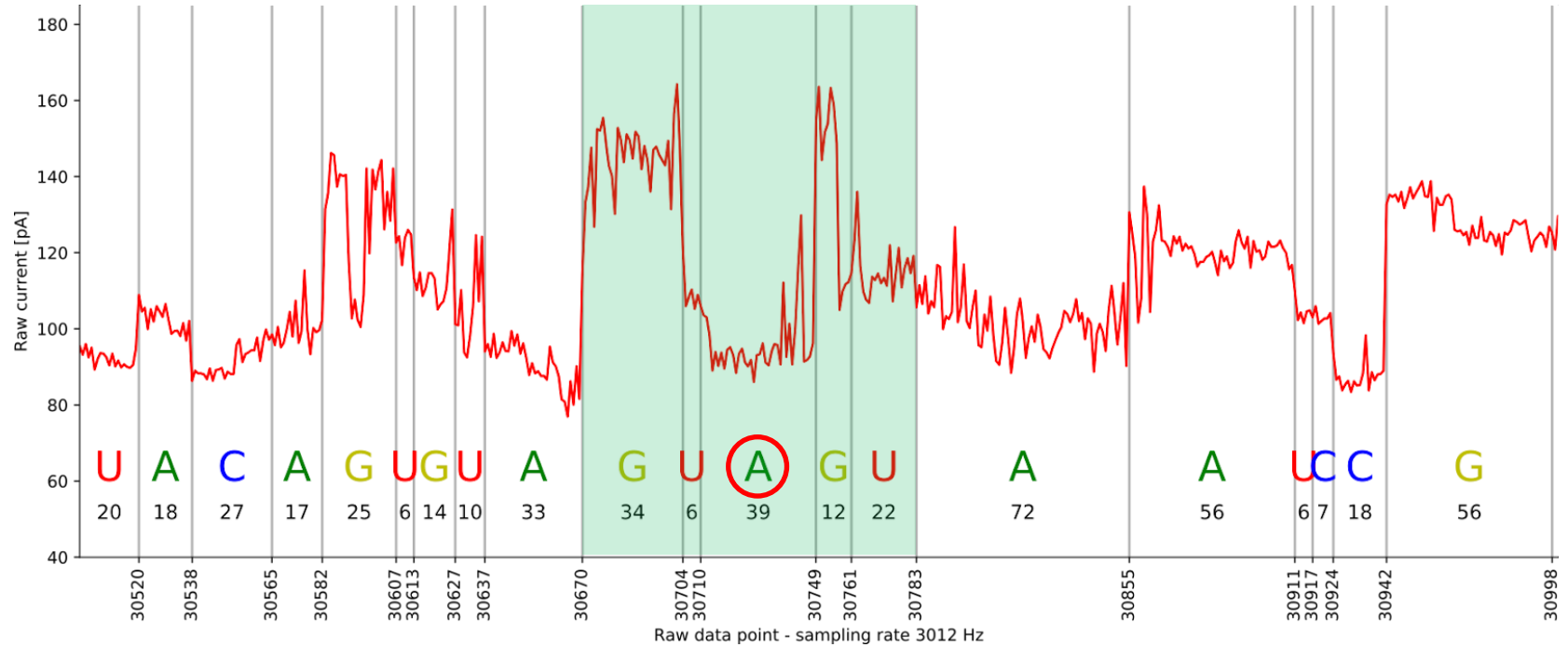




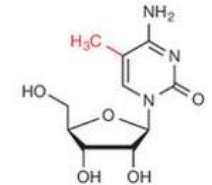
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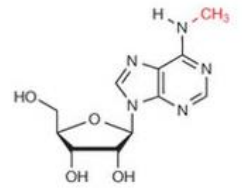
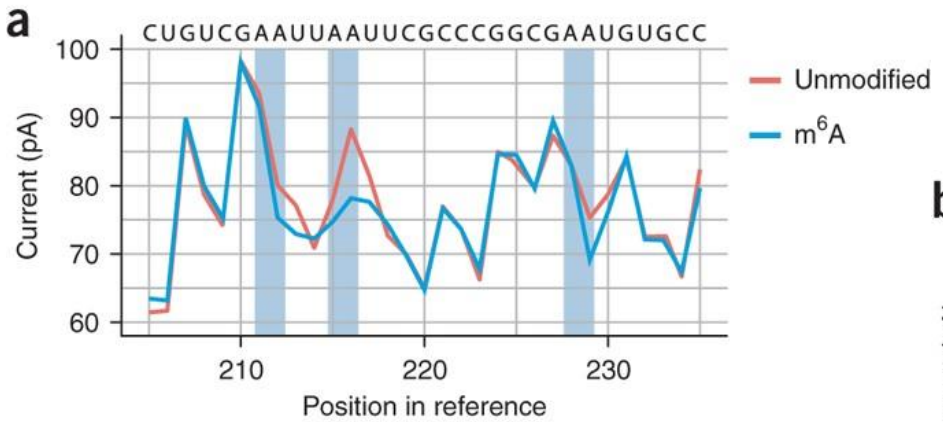
# Raw signal of a RNA



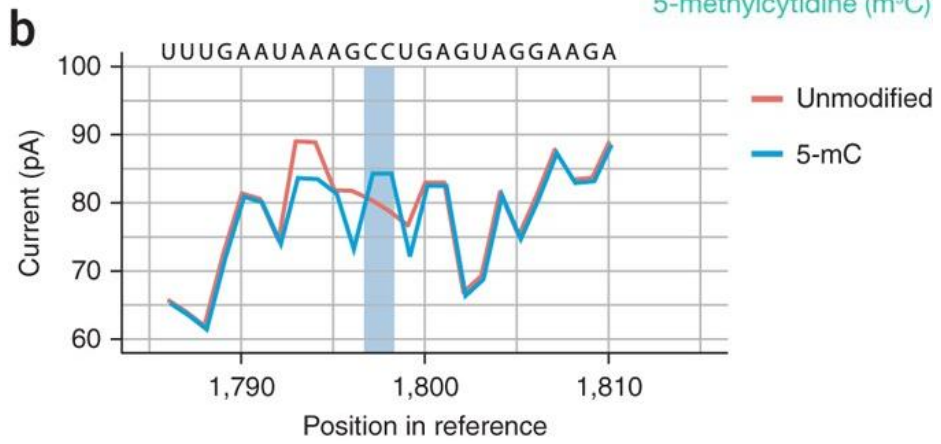
# RNA modifications



5-methylcytidine (m<sup>5</sup>C)



N<sup>6</sup>-methyladenosine (m<sup>6</sup>A)



Garalde, D., Snell, E., Jachimowicz, D. *et al.* Highly parallel direct RNA sequencing on an array of nanopores. *Nat Methods* 15, 201–206 (2018)



# Creating training data for m6A detection via *in vitro* transcription (IVT)

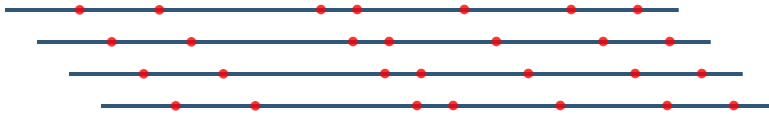


Known dsDNA template



## m6A modified reads

- Transcribe with T7
- m6A, C, G, U



## unmodified reads

- Transcribe with T7
- A, C, G, U

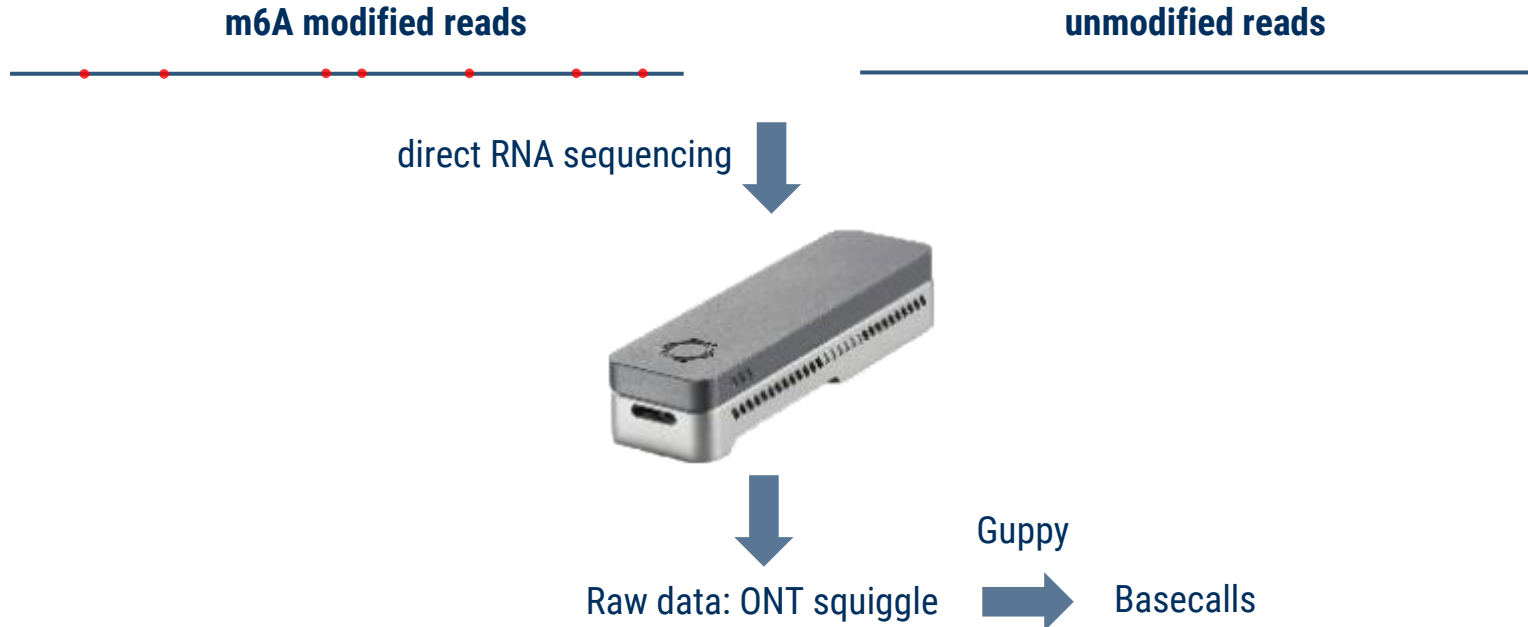


Data design from: Liu, H., Begik, O., Lucas, M.C. *et al.* Accurate detection of m6A RNA modifications in native RNA sequences. Nat Commun 10, 4079 (2019)





# Creating training data for m6A detection via *in vitro* transcription (IVT)

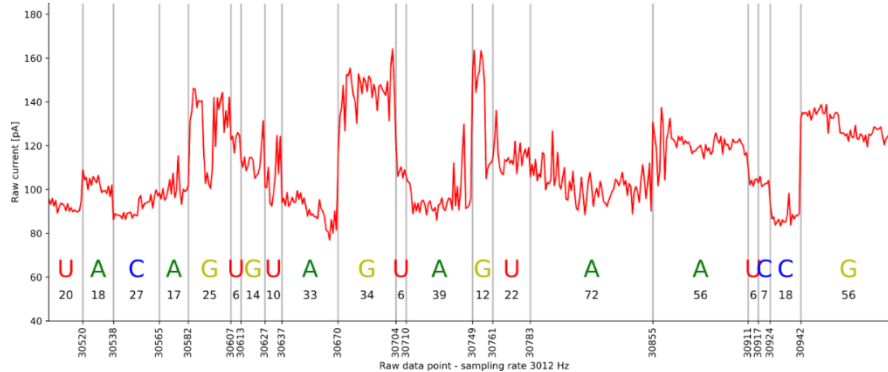


# Creating training data for m6A detection via *in vitro* transcription (IVT)

- Raw data: ONT squiggle
- Basecalled reads
- Reference sequence
- Mapping



Resquigging with **nanopolish eventalign**  
(basecalling error correction and signal segmentation)



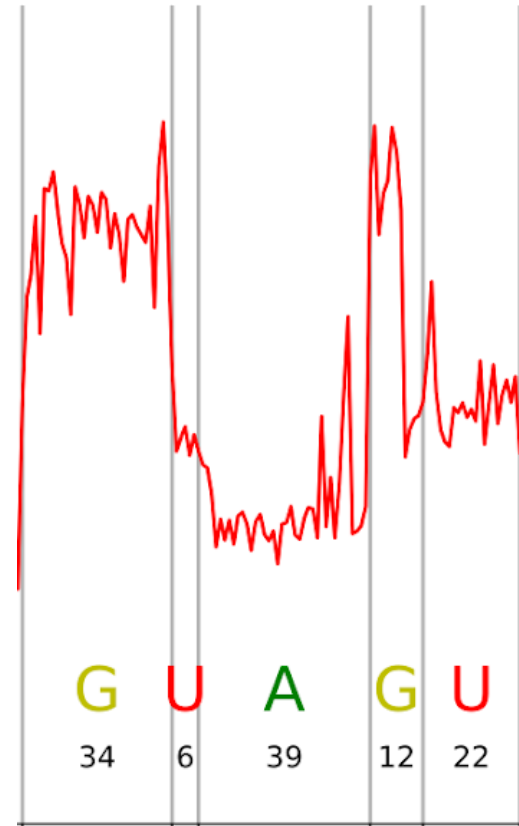
Loman, N., Quick, J. & Simpson, J. A complete bacterial genome assembled de novo using only nanopore sequencing data. Nat Methods 12, 733–735 (2015).



# What is the input?

Sample: 5mer containing a m6A or canonical A in the middle

- Features:
  - Corrected basecalls
    - Use an embedding layer to encode the bases (A, C, G, U)
    - m6A and A will have the same encoding
  - Segmented signal
    - Extract the segments and interpolate them to a given size
  - Segment size
  - Trace from Guppy (base transition probabilities, bad resolution, 1 trace every 10<sup>th</sup> measured datapoint)



## Results on datasets

- Train on IVT dataset from Liu *et al.*
- Test on our IVT dataset (different dataset, similar design)
- Test on *in vivo* dataset from Göke *et al.*
  - very bad m6A detection
  - Not transferable to *in vivo* data...?

Dataset	Samples #	Acc.
Training on IVT of Liu <i>et al.</i> (80:20 split)	4'888'798 <ul style="list-style-type: none"><li>• Mod: 2'444'399</li><li>• Can: 2'444'399</li></ul>	0.95 on 20% split
Testing on IVT of Manja <i>et al.</i>	1'394'076 <ul style="list-style-type: none"><li>• Mod: 697'038</li><li>• Can: 697'038</li></ul>	0.73
Testing for TP on <i>in vivo</i> data from Göke <i>et al.</i>	Mod: 1'252'679	0.25



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# Challenges

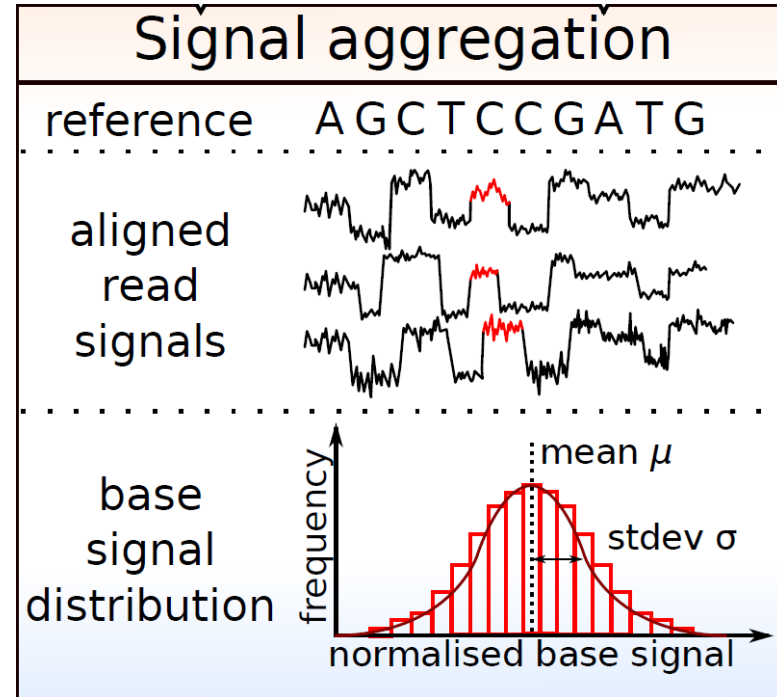
- How can we design the *in vitro* transcription experiments as natural as possible and still know which positions are modified?
- Where/How can we get ground truth for *in vivo* data for modifications?
- Do you know or have ONT data with modifications and have a ground truth that we could use?
- Which input features should be used and how should they be provided/feeder to the model?





## Magnipore (not published yet)

- Compares two samples sequenced with ONT
- Collect signals per reference positions from reads to calculate signal distributions
- Compare these signal distributions between the samples per position
- Look for significant signal differences
- Differences can originate from mutations or modifications





## Isotopic labeling with D2O

- Detect deuterium labeled nucleotide sequences with ONT
- Isotopes are much smaller modifications
- Currently we see minor changes between H2O and D2O
- The signal-to-noise ratio is currently too small for accurate detection



Thanks to:

- Manja Marz
  - Christian Höner zu Siederdisen
  - Sebastian Krautwurst
  
  - Wetlab:
    - Akash Srivastava
    - Milena Žarković
- and you!

Thanks for your attention!

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**DFG** Deutsche  
Forschungsgemeinschaft  
German Research Foundation



Thüringer Zentrum für  
Lernende Systeme und Robotik

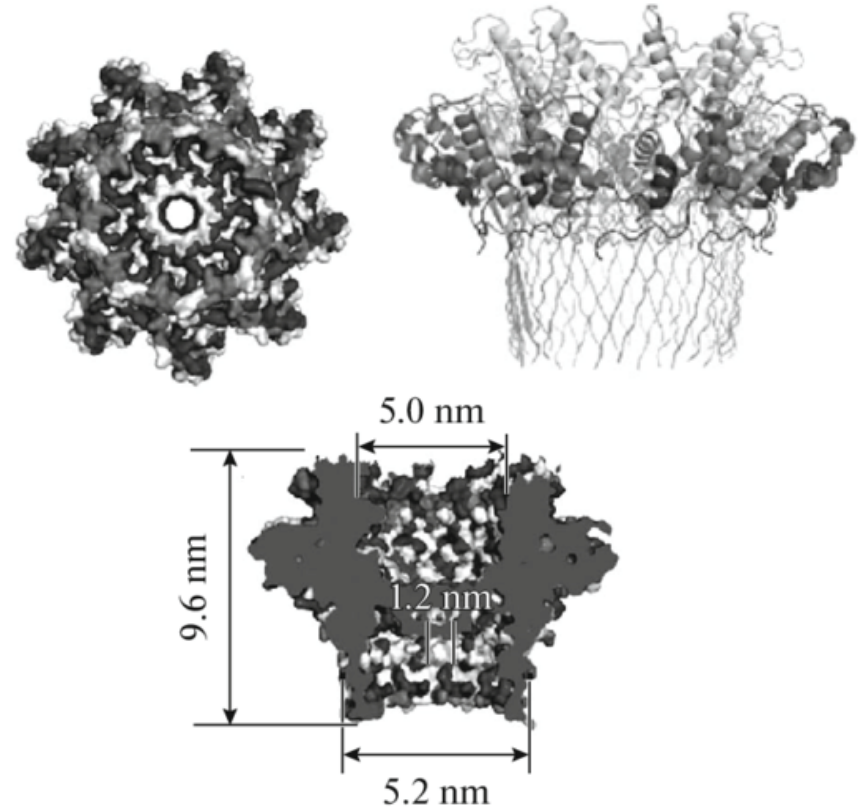


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- German DFG Collaborative Research Centre AquaDiva (CRC 1076 AquaDiva)
- The German state of Thuringia via the Thüringer Aufbaubank (2021 FGI 0009)

# What is a nanopore?

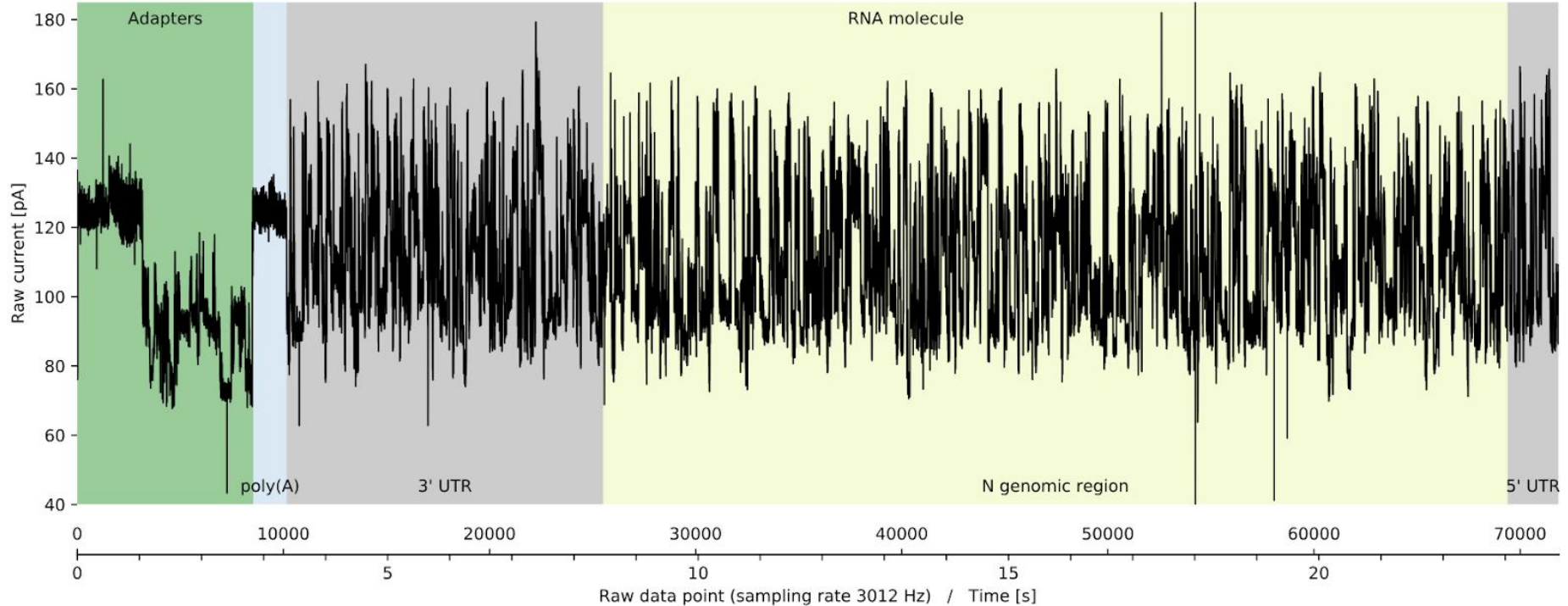
- Voltage is applied to the membrane
- Measured current is characteristic to the molecules at the most narrow part of the pore
- 5 nucleotides are measured at once
- Measurements are influenced by the molecules in the pore, the pore, the sensor, the flowcell and the sequencing kit/protocol



Barkova, D.V., Andrianova, M.S., Komarova, N.V. *et al.* Channel and Motor Proteins for Translocation of Nucleic Acids in Nanopore Sequencing. *Moscow Univ. Chem. Bull.* 75, 149–161 (2020).



# Raw signal of a RNA

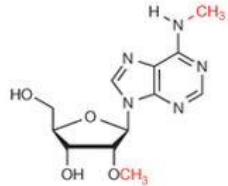


Viehweger *et al.*, Direct RNA nanopore sequencing of full-length coronavirus genomes provides novel insights into structural variants and enables modification analysis, Genome Research 2019

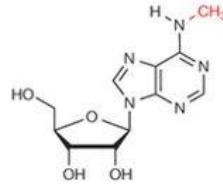




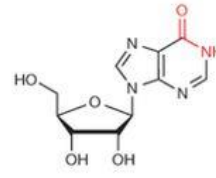
# RNA modifications



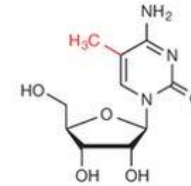
*N*<sup>6</sup>,2'-O-dimethyladenosine (m<sup>6</sup>Am)



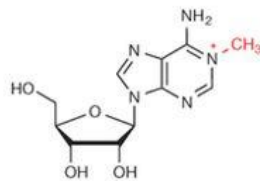
*N*<sup>6</sup>-methyladenosine (m<sup>6</sup>A)



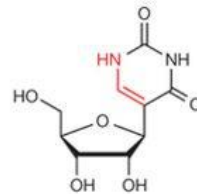
Inosine (I)



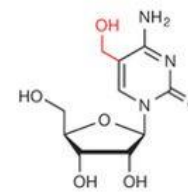
5-methylcytidine (m<sup>5</sup>C)



*N*<sup>1</sup>-methyladenosine (m<sup>1</sup>A)



Pseudouridine (Ψ)



5-hydroxymethylcytidine (hm<sup>5</sup>C)



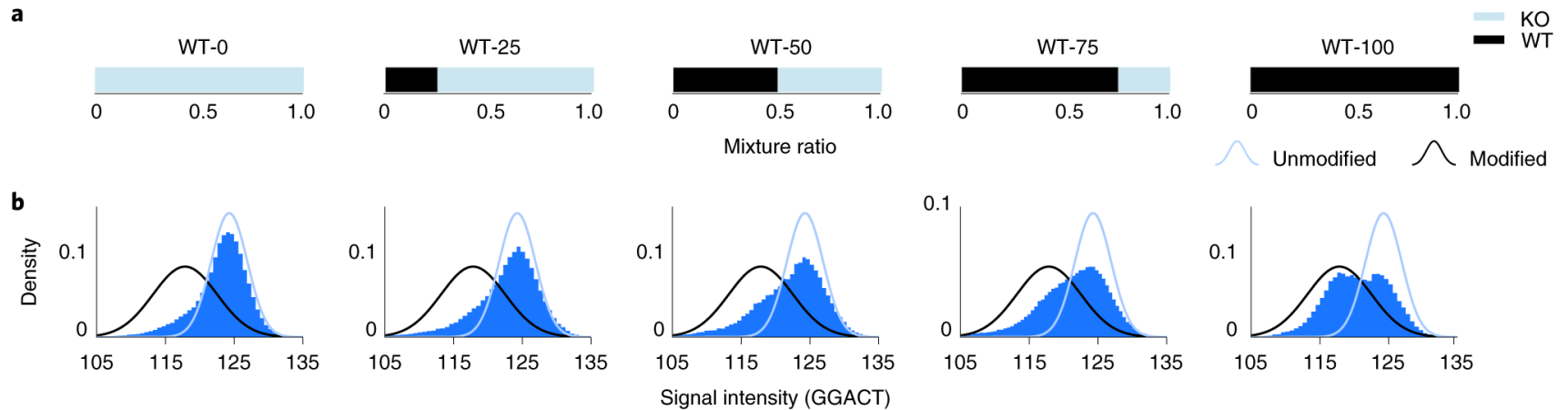
# Signal normalisation

- Make the sequencing comparable across different
  - Pores/sensors
  - Flowcells
  - Sequencing protocols/experiments

$$\textit{normalised signal} = \frac{\textit{signal} - \textit{median}(\textit{signal})}{\textit{median\_absolute\_deviation}(\textit{signal})}$$

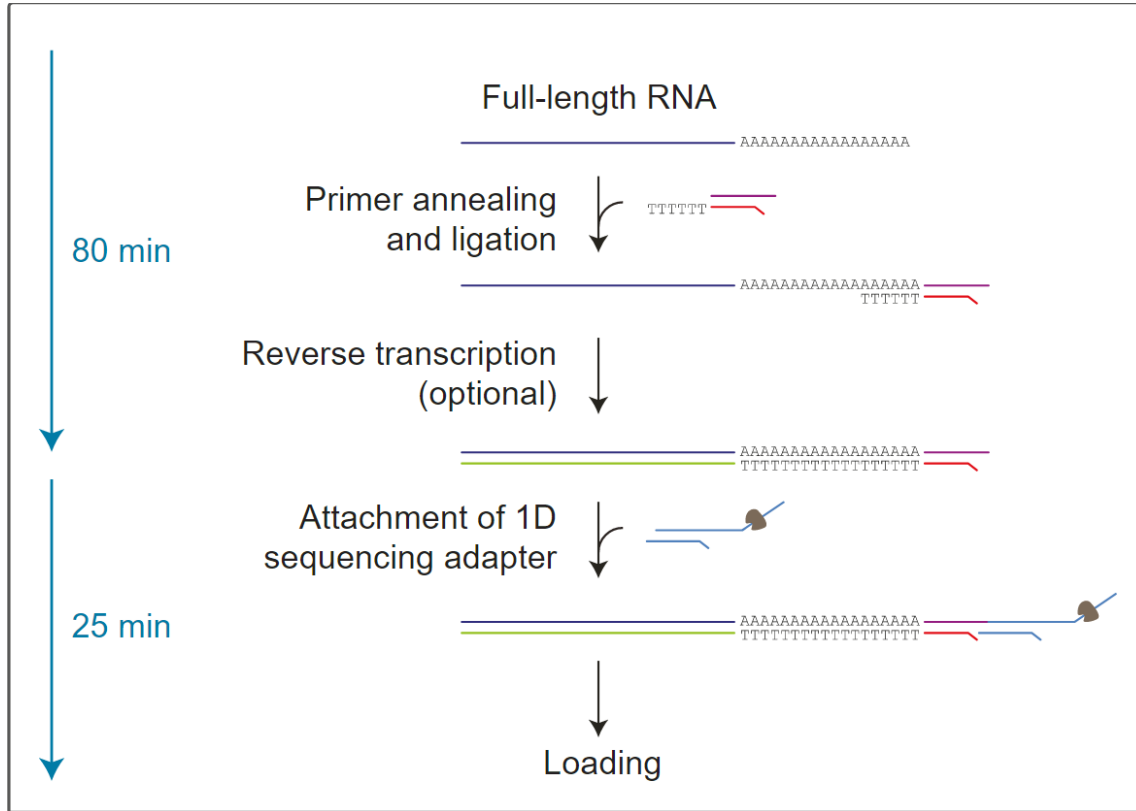


# RNA modifications



Pratanwanich, P.N., Yao, F., Chen, Y. et al. Identification of differential RNA modifications from nanopore direct RNA sequencing with xPore. *Nat Biotechnol* 39, 1394–1402 (2021)





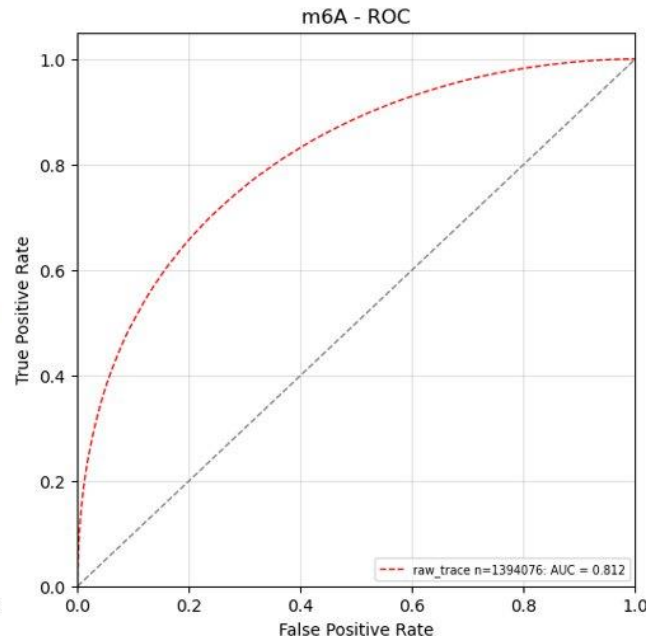
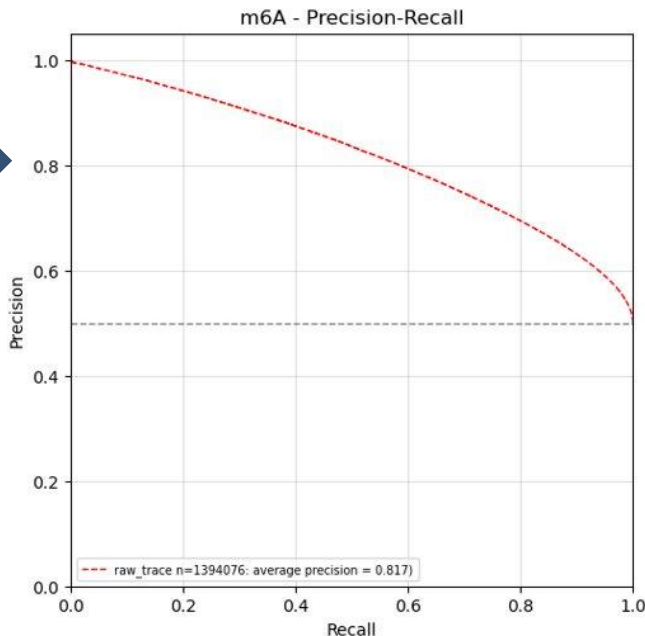
# Results

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- Test on our IVT dataset (different dataset, similar design)
- Test on in vivo dataset from Goeke et. al.
  - very bad m6A detection
  - Not transferable to in vivo data...?



- $Precision = \frac{TP}{TP+FP}$
- $Recall = \frac{TP}{TP+FN}$

- $True\ positive\ rate = recall$
- $False\ positive\ rate = \frac{FP}{FP+TN}$





## Datasets

IVT datasets	Liu <i>et al.</i>	Manja <i>et al.</i>
# Reads	88,819	39,219
# Canonical	47,325	9,542
# Modified	11,921	26,337

<i>In vivo</i> datasets by Göke <i>et al.</i>	Wild Type			Knockout		
	1	2	3	1	2	3
# Reads	2,389,434	3,302,095	1,124,426	3,476,668	4,265,961	3,993,818



# Neural Network

- Embedding layer for the base encoding
- Transformer layer for the signal processing
- Linear fully connected layers
- One output value from a sigmoid function predicting the modification status for the A in the input sample
  - Output  $< 0.5$  = unmodified
  - Output  $\geq 0.5$  = modified

