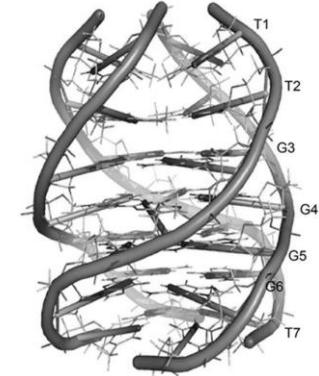


CRAIG WANTS ME TO TALK ABOUT G4S

MARTA SZACHNIUK

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RNA Benasque, 18.08.2022



Institute of Computing Science,
Poznan University of Technology,
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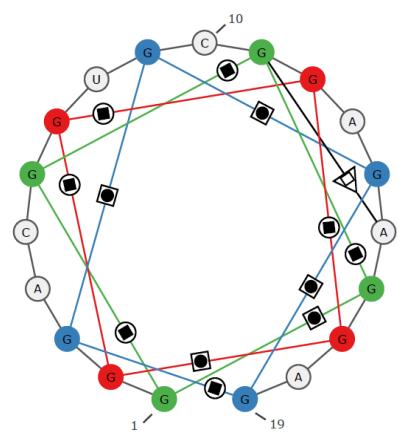


Institute of Bioorganic Chemistry
Polish Academy of Sciences,
Poznan, Poland

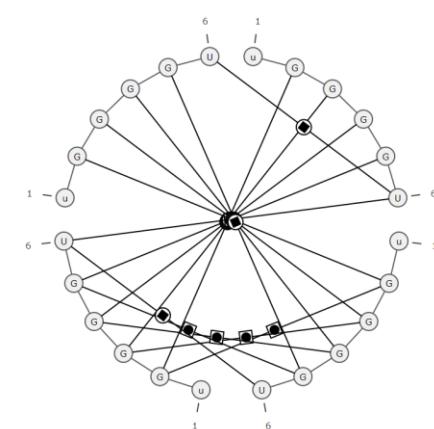
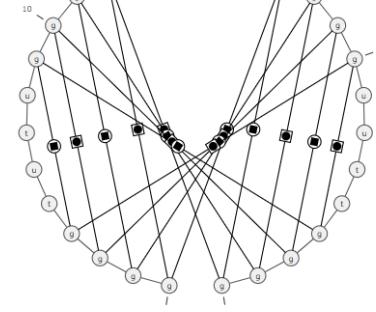
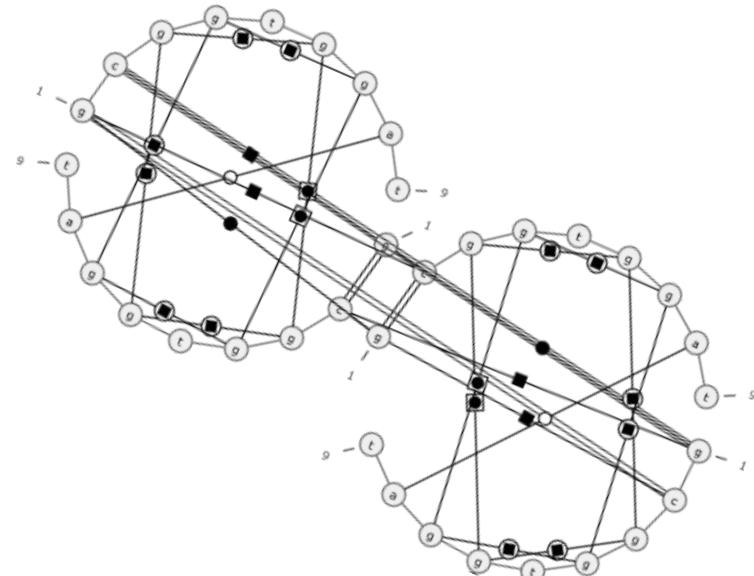
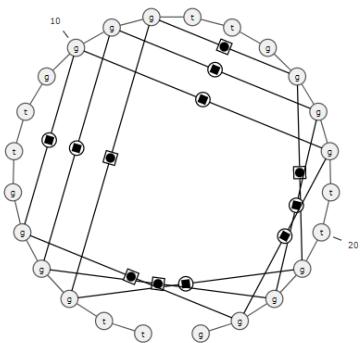
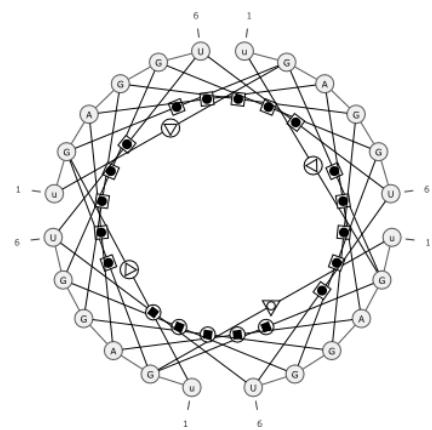
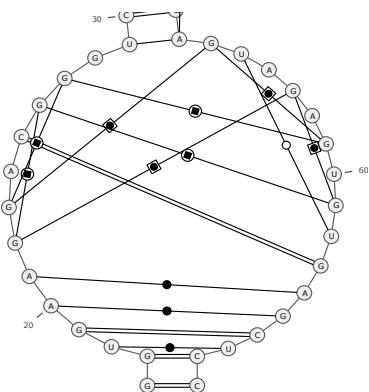




November 19, 2016



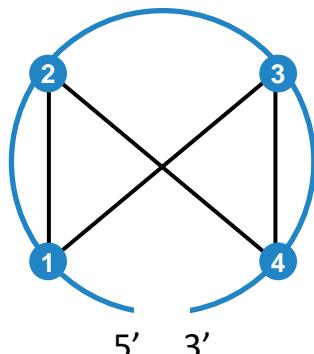
- Hoogsteen - Watson-Crick
- Watson-Crick - Hoogsteen



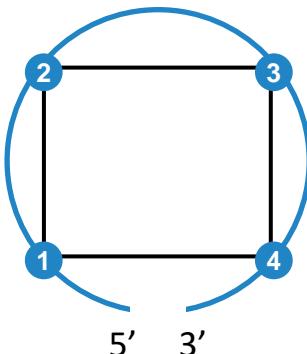
2D STRUCTURE DIAGRAMS → ONZ CLASSIFICATION

cyclic pairing schemes

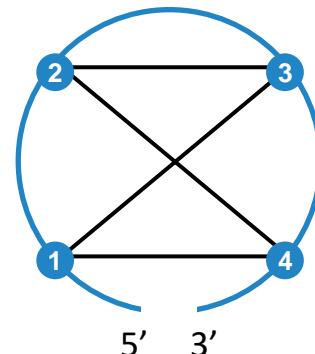
N



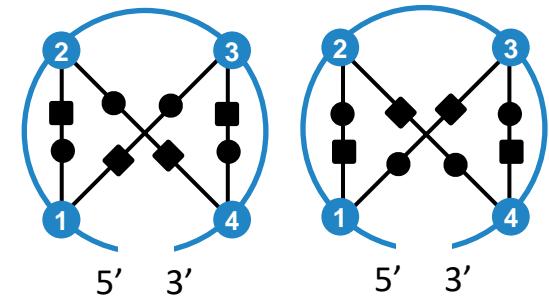
O



Z

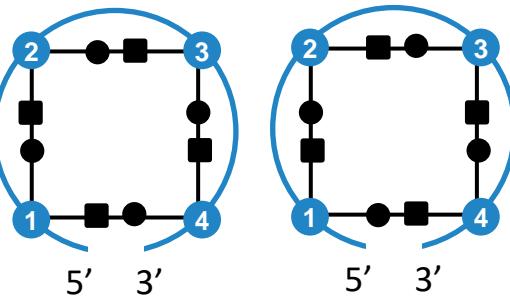


- Watson-Crick
- Hoogsteen



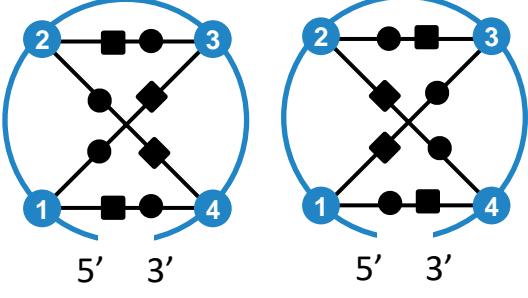
N+

N-



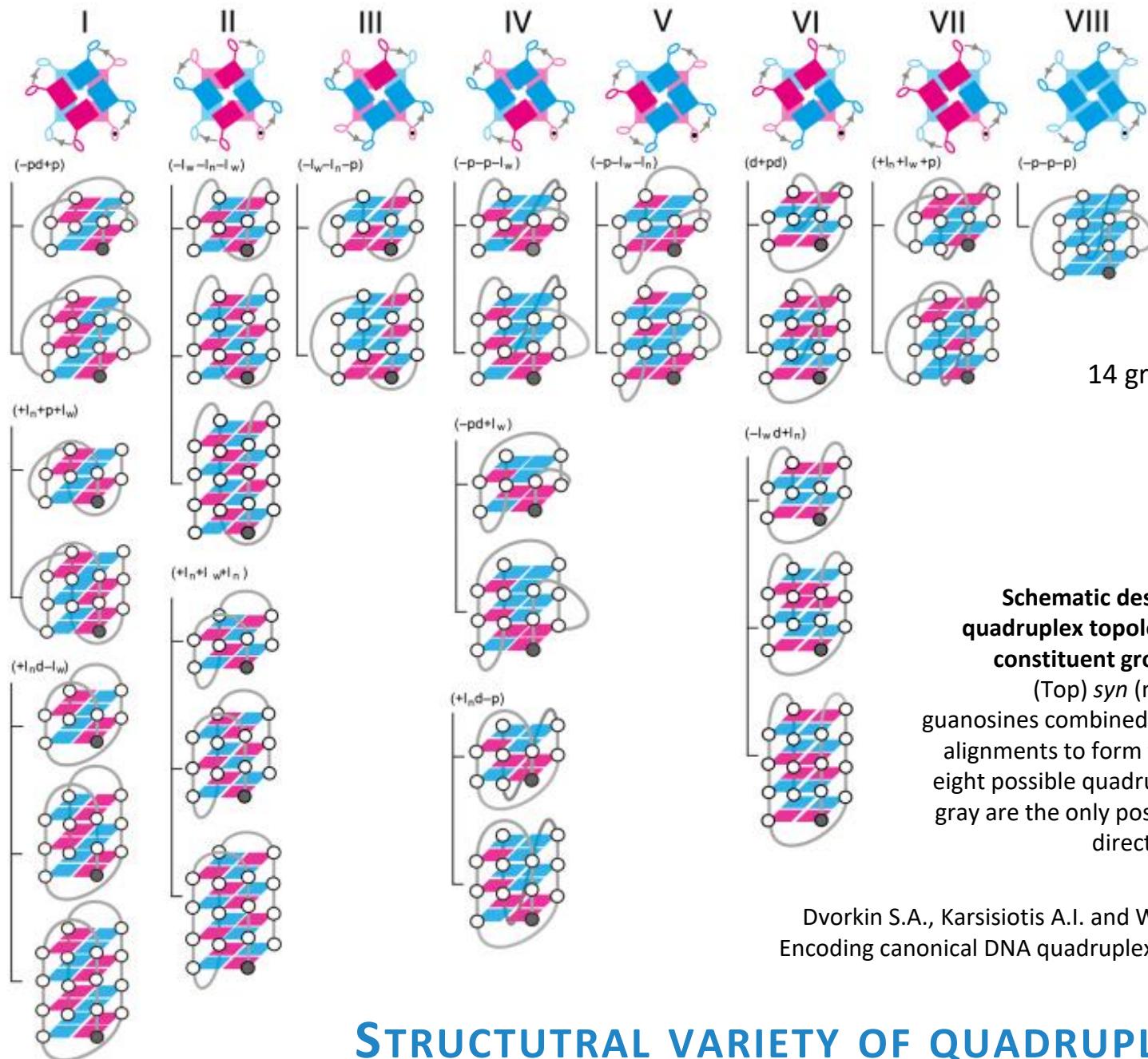
O+

O-



Z+

Z-



Schematic description of all canonical quadruplex topologies feasible with their constituent groove type combination.

(Top) *syn* (magenta) and *anti* (cyan) guanosines combined through hydrogen bond alignments to form tetrads representing the eight possible quadruplex stems. Indicated in gray are the only possible strand progression directionalities for each stem.

Dvorkin S.A., Karsiotis A.I. and Webba da Silva M. (2018)
Encoding canonical DNA quadruplex structure. Sci Adv, 4(8).

STRUCTURAL VARIETY OF QUADRUPLEXES

DESCRIBING QUADRUPLEX STRUCTURE

- Loops (lateral, diagonal, propeller)
- Parallel/antiparallel directionality
- Right-handed/left-handed torsion
- Number of tetrads
- Sequence
- G-tracts
- Number of strands
- Rise and twist parameters
- Groove
- Planarity deviation
- Webba da Silva Classification
- ONZ classification (by our group)
- and more...

Zok et al. BMC Bioinformatics (2020) 21:40
<https://doi.org/10.1186/s12859-020-3385-1>

BMC Bioinformatics

SOFTWARE

Open Access

EITetrad: a tool for identification and classification of tetrads and quadruplexes

Tomasz Zok^{1,2}, Mariusz Popenda³ and Marta Szachniuk^{1,3*} 



EITetrad: a tool for identification and classification of tetrads and quadruplexes

Abstract Quadruplexes are unique nucleic acid structures, e.g., hairpins and intermolecular quadruplexes. To study these motifs, we focused mainly on the sequence and the three strands are used. In this paper, we present EITetrad, a software that identifies specific features of quadruplexes and classifies them into four groups: parallel, antiparallel, lateral, and diagonal. The results are presented in two ways: by the sequence or by the structure. The structure is shown as a 3D model. The results are also presented in a tabular form. The software is available at www.eitetrad.com.

Keywords: Quadruplex, Classification, Secondary structure mapping, One-letter notation, 3D diagram

Background Quadruplexes are unique nucleic acid structures, e.g., hairpins and intermolecular quadruplexes. To study these motifs, we focused mainly on the sequence and the three strands are used. In this paper, we present EITetrad, a software that identifies specific features of quadruplexes and classifies them into four groups: parallel, antiparallel, lateral, and diagonal. The results are presented in two ways: by the sequence or by the structure. The structure is shown as a 3D model. The results are also presented in a tabular form. The software is available at www.eitetrad.com.

Results Here, we present EITetrad, a software that identifies specific features of quadruplexes and classifies them into four groups: parallel, antiparallel, lateral, and diagonal. The results are presented in two ways: by the sequence or by the structure. The structure is shown as a 3D model. The results are also presented in a tabular form. The software is available at www.eitetrad.com.

Conclusion EITetrad is a tool for identification and classification of tetrads and quadruplexes. It supports the analysis of parallel and antiparallel motifs. Results are now accessible online. The software is freely available at www.eitetrad.com.

Availability The software is freely available at www.eitetrad.com. The source code is available at <https://github.com/tzok/EITetrad>.

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BMC Bioinformatics

Software

Open Access

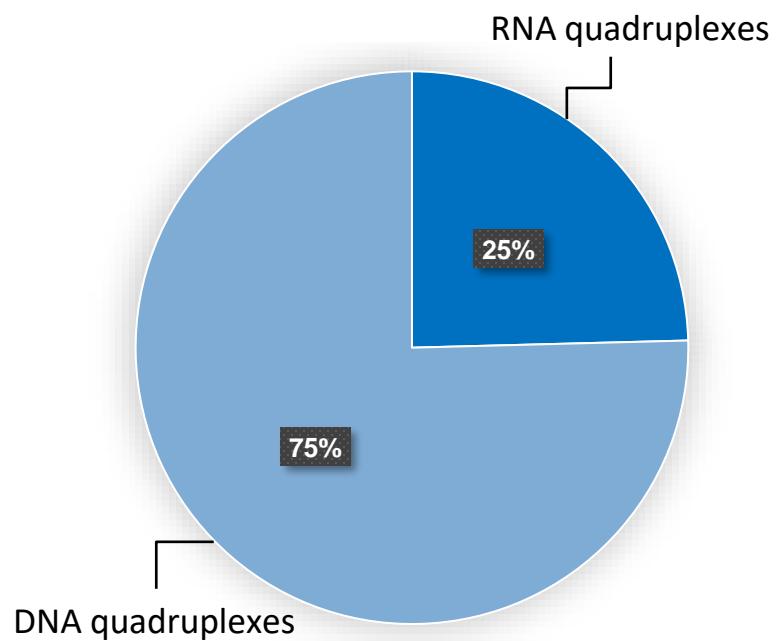
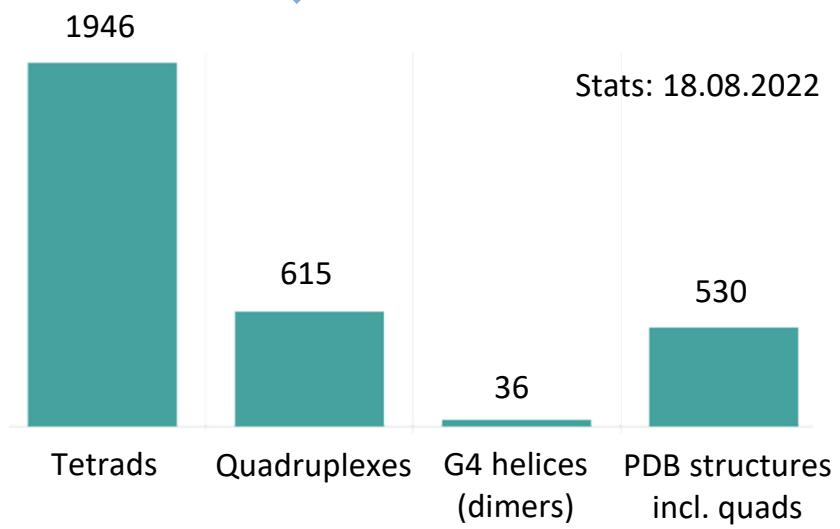
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RG4 AND ITS 2D/3D STRUCTURE PROCESSING

	classification		computation of structure parameters						visualization		
	G4 Classification	Base-pair Classification	Strand polarity	Area	Rise	Twist	Planarity	Torsion angles	2D view	3D view	Moving camera
DSSR	Loop-based	Saenger, LW	+	+	+	+	+	+	+	+	+
ElTetrad	ONZ	LW	+		+	+	+			+	
RNApdbee		Saenger, LW								+	
3D-Nus	Q1-Q17				+	+		+		+	+

- DSSR <http://skmatic.x3dna.org/> LW: Leontis-Westhof classification
- ElTetrad <https://github.com/tzok/eltetrad/>
- RNApdbee <http://rnapdbee.cs.put.poznan.pl/>
- 3D-Nus <http://iith.ac.in/3dnus/>



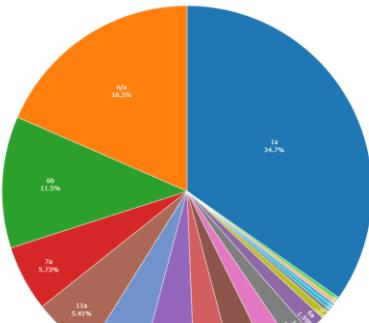
structure parameters

Tetrad pairs	Twist [°]	Rise [\AA]	Strand direction
T26 - T27	18.46	3.37	antiparallel
T27 - T28	35.82	3.26	antiparallel
T28 - T29	21.54	3.37	antiparallel

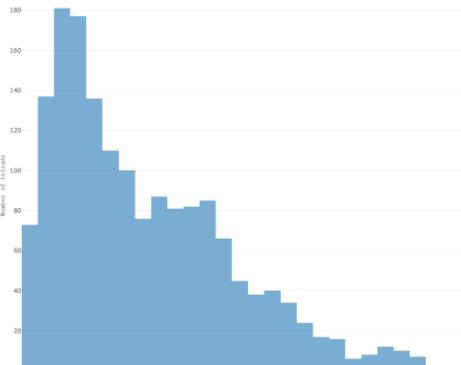
Loop ID	Short sequence	Full sequence	Loop length	Loop type
L1	TTTT	A.DT5, A.DT6, A.DT7, A.DT8	4	diagonal
L2	TTTT	B.DT17, B.DT18, B.DT19, B.DT20	4	diagonal

Tetrad ID	Nucleotide 1	Nucleotide 2	Nucleotide 3	Nucleotide 4
T26	60.24° / syn	56.09° / syn	-115.09° / anti	-108.173° / anti
T27	-114.73° / anti	62.16° / syn	43.97° / syn	-124.417° / anti
T28	62.32° / syn	44.03° / syn	-124.21° / anti	-114.899° / anti
T29	-108.05° / anti	60.19° / syn	55.98° / syn	-115.168° / anti

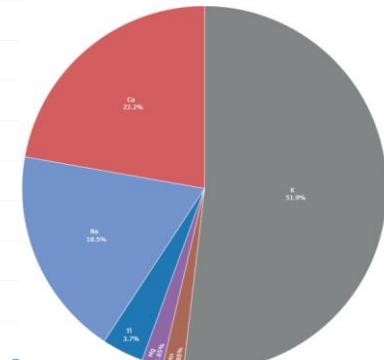
statistics



Webba da Silva classes



planarity

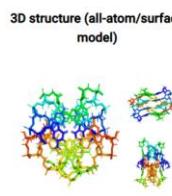
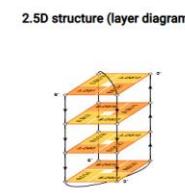
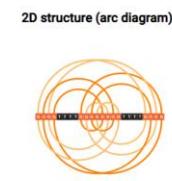
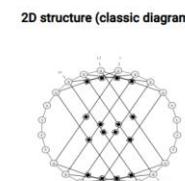


ions

structure visualizations, dot-brackets

2D structure (dot-bracket):

GGGGTTTGCCCC-GGGGTTTGCCCC
(((((....(((([-))))....))))))
(((....(((([-))))....))))))



OTHER QUADRUPLEX-RELATED DATABASES (16)

	DNA	RNA	G4 verification	G4 sequence	No. of G4s	DB records	Customized search	Web interface	Visual output
DSSR-G4DB	+	+	Experimental	+	354	354 (PDBs)	+	+	+
G4IPDB	+	+	Experimental	+	unknown	216	+	+	+
G4LDB	+	+	Experimental	+	unknown	>800	+	+	+
G4RNA	+	+	Experimental	+	321	567	+	+	
Lit392	+	+	Experimental	+	298	392			
Lit638	+	+	Experimental	+	506	638			
GSE63874	+		Experimental		716 310	32 mln			
GSE77282		+	Experimental		3 383	1,15 bln			
GSE110582	+		Experimental		1 420 841	767,39 Mb			
GSE129281		+	Experimental		329	3 505			
Greglist	+		In silico	+	Unknown	115 442	+	+	+
GRSDB2		+	In silico	+	3 255 075	29 288	+	+	+
G4-virus	+	+	In silico		47	248		+	+
Non-B DB	+		In silico	+	3 864 596	12	+	+	+
PlantGQ	+		In silico	+	626 341 645	195	+	+	+
Quadbase2	+		In silico	+	unknown	1 897	+	+	+

JOURNAL ARTICLE

How bioinformatics resources work with G4 RNAs 

Joanna Miskiewicz, Joanna Sarzynska, Marta Szachniuk

Briefings in Bioinformatics, Volume 22, Issue 3, May 2021, bbba201, <https://doi.org/10.1093/bib/bbba201>

Published: 08 September 2020 Article history ▾

Sequence analysis and prediction of sites where quadruplexes can form (14)

- G4Hunter
 - G4-iM Grinder
 - G4Predict
 - G4RNA Screener
 - G4Catchall
 - G4-Predictor
 - Pqsfinder
 - QGRS Mapper
 - G4PromFinder
 - ImGQfinder
 - Quadron
 - QPARSE
 - G4P Calculator
 - TetraplexFinder

Secondary structure prediction (1)

- RNAfold

Analysis of 2D & 3D structure (4)

- DSSR
 - RNApdbee
 - ElTetrad
 - 3D-Nus

Databases (16)

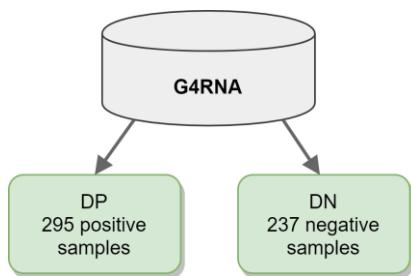
- G4LDB
 - G4-virus
 - DSSR-G4DB
 - Quadbase2
 - GSE63874
 - G4RNA
 - GRSDB2
 - Lit392
 - Plant-GG
 - GSE77282
 - G4IPDB
 - Greglist
 - Lit638
 - Non-B DB
 - GSE110582
 - GSF129281

G4 PATTERNS IN PQS PREDICTION ALGORITHMS

Tool	Pattern	Pattern description	Exemplary motif	
G4Catchall	G_n	Perfect G-tract	GGG	$n = 3$
	$\text{G}_x \text{N}_y \text{G}_{(n-x)}$	Bulged G-tract	GGAG	$n = 3; x = 2; y = 1$
	$\text{G}_x \text{N}_y \text{G}_{(n-x-1)}$	Mismatched G-tract	GGAG	$n = 4; x = 2; y = 1$
G4Hunter	n/a	n/a	n/a	
G4-iM Grinder	n/a	n/a	n/a	
G4P Calculator	G_n	Perfect G-tract	GGG	$n = 3$
G4Predict (Intra)	$\text{G}_{x1} \text{N}_{y1} \text{G}_{x2} \text{N}_{y2} \text{G}_{x3} \text{N}_{y3} \text{G}_{x4}$	General pattern for intramolecular PQS	GGAAAGGAAAGGAAGG	$x1 = x2 = x3 = x4 = 2; y1 = y3 = 2; y2 = 3$
G4-Predictor V.2	$\text{G}_{2-6} \text{N}_{0-36} \text{G}_{2-6} \text{N}_{0-36} \text{G}_{2-6} \text{N}_{0-36} \text{G}_{2-6}$	Patterns depend on users' settings	GGAGGGAAAGGAAGGG	
G4PromFinder	$\text{G}_{x1} \text{N}_{y1} \text{G}_{x2} \text{N}_{y2} \text{G}_{x3} \text{N}_{y3} \text{G}_{x4}$	General pattern for PQS	GGAGGGAAAGGAAGGG	$x1 = x3 = 2; x2 = x4 = 3; y1 = 1; y2 = y3 = 2$
G4RNA screener	n/a (ML algorithm)	n/a	n/a	
ImGQfinder	G_n	Perfect G-tract	GGG	$n = 3$
	$\text{G}_{i-1} \text{N} \text{G}_{n-i+1}$	Bulged G-tract	GGAGG	$n = 4; i = 3$
	$\text{G}_{i-1} \text{N} \text{G}_{n-i}$	Mismatched G-tract	GGAG	$n = 4; i = 3$
pqsfinder	$\text{G}_{x1} \text{N}_{y1} \text{G}_{x2} \text{N}_{y2} \text{G}_{x3} \text{N}_{y3} \text{G}_{x4}$	General pattern for PQS	GGAGGAAGGAAGGG	$x1 = x2 = x3 = 2; x4 = 3; y1 = 1; y2 = y3 = 2$
QGRS Mapper	$\text{G}_{x1} \text{N}_{y1} \text{G}_{x2} \text{N}_{y2} \text{G}_{x3} \text{N}_{y3} \text{G}_{x4}$	General pattern for PQS	GGAGGAAGGAAGG	$x1 = x2 = x3 = x4 = 2; y1 = 1; y2 = y3 = 2$
QPARSE	n/a	Patterns depend on users' settings	n/a	
Quadron	$\text{G}_{x1} \text{N}_{y1} \text{G}_{x2} \text{N}_{y2} \text{G}_{x3} \text{N}_{y3} \text{G}_{x4}$	General pattern for PQS	GGAGGAAGGAAGG	$x1 = x2 = x3 = x4 = 2; y1 = 1; y2 = y3 = 2$
TetraplexFinder	$\text{G}_{x1} \text{N}_{y1} \text{G}_{x2} \text{N}_{y2} \text{G}_{x3} \text{N}_{y3} \text{G}_{x4}$	General pattern for PQS	GGAAAGGAAGGAAGG	$x1 = x2 = x3 = x4 = 2; y1 = 3; y2 = y3 = 2$

BENCHMARKING ALGORITHMS FOR G4 LOCATION PREDICTION

tested for RNA quadruplexes



Positive (DP) and negative (DN) datasets

Best results:
G4RNA screener
G4Catchall
RNAfold

Coverage of DP and DN datasets with correct predictions: positive in DP and negative in DN [%].



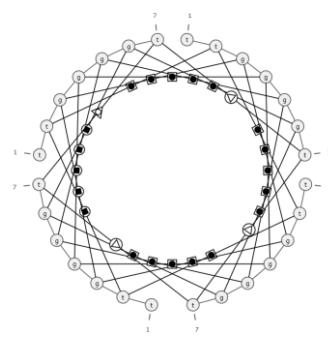
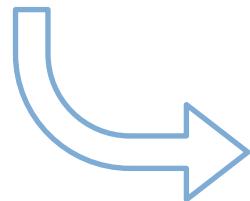
BIG CHALLENGES - PREDICTION

TTGGGGTAACACCTTG
GGGTGCCTGGGGTAT
ATACCCCCCTTGGGG



TTGGGGTAACACCTTG**
GGGTGCCTGGGGTAT**
ATACCCCCC**TTGGGG******

- Predict the location of non-G-quads
- Predict the secondary structure
(non-canonical bps!)
- Predict the 3D structure of quadruplexes





Tomasz Żok



Marta Szachniuk



Joanna Miśkiewicz



Michał Żurkowski



Natalia Kraszewska



Mariusz Popenda



Joanna Sarzyńska

Take home message ☺

